

REVIEW ARTICLE

Obesogens, stem cells and the developmental programming of obesity

A. Janesick* and B. Blumberg*,†

Departments of *Developmental and Cell Biology and †Pharmaceutical Sciences, University of California, Irvine, CA, USA

Summary

Keywords:

development, endocrine disrupters, environmental factors, epigenetics, fish < animal models, hormone receptors, mouse < animal models, obesity, phthalates, sex hormones

Correspondence:

Bruce Blumberg, Department of Developmental and Cell Biology, 2011 BioSci 3, University of California, Irvine, CA 92697-2300, USA. E-mail: blumberg@uci.edu

Received 1 September 2011; revised 28 December 2011; accepted 4 January 2012

doi:10.1111/j.1365-2605.2012.01247.x

Obesogens are chemicals that directly or indirectly lead to increased fat accumulation and obesity. Obesogens have the potential to disrupt multiple metabolic signalling pathways in the developing organism that can result in permanent changes in adult physiology. Prenatal or perinatal exposure to obesogenic endocrine disrupting chemicals has been shown to predispose an organism to store more fat from the beginning of its life. For example, excess oestrogen or cortisol exposure in the womb or during early life resulted in an increased susceptibility to obesity and metabolic syndrome later in life. This review focuses on the effects of environmental chemicals, such as the model obesogen, tributyltin (TBT), on the development of obesity. We discuss evidence linking the obesogenic effects of TBT with its ability to activate the peroxisome proliferator-activated receptor gamma and stimulate adipogenesis. We also discuss how TBT and other environmental obesogens may lead to epigenetic changes that predispose exposed individuals to subsequent weight gain and obesity. This suggests that humans, who have been exposed to obesogenic chemicals during sensitive windows of development, might be pre-programmed to store increased amounts of fat, resulting in a lifelong struggle to maintain a healthy weight and exacerbating the deleterious effects of poor diet and inadequate exercise.

Introduction

We are in the midst of a worldwide obesity epidemic that is particularly apparent in the United States. Currently, 34% of American adults are obese (body mass index > 30) and an additional 68% are overweight (BMI > 25), double the worldwide average and 10 times the rates in Korea and Japan (Flegal *et al.*, 2010). The fraction of overweight US adults is predicted to increase to 86% by 2020 (Flegal *et al.*, 2010). BMI is a simple measurement that does not distinguish among increased subcutaneous adiposity, which is generally considered to be the preferred storage depot for excess calories (Ibrahim, 2010), increased visceral adiposity, which is a pathological condition that increases the risk of cardiovascular disease, metabolic syndrome and diabetes (Freedland, 2004) and increased muscularity, which is not a risk factor for metabolic diseases. However, the trend towards higher BMI in the population is largely accompanied by

increased visceral adiposity and associated metabolic syndrome disorders (Lustig, 2006).

The most typical explanation given for the increased rate of obesity is that consumption of calorie dense food has increased and that physical activity has decreased, the thermodynamic or 'calories in-calories out' model. Obviously, to gain weight, calories consumed must exceed calories burned; however, the situation is not as simple as balancing one's caloric checkbook. The biochemical nature of the calories consumed plays a very large role in how and where they are stored, as well as in the regulation of appetite and satiety (Lustig, 2006, 2010). In addition, there are considerable differences in how individuals accrue weight given the same amount of excess calories. Once an individual becomes obese, it is difficult to lose weight and sustain weight loss due to highly efficient homeostatic mechanisms regulating energy balance (Butte *et al.*, 2007; Muhlhausler & Smith, 2009).

The observation that people do not accrue weight equally given the same amount of caloric excess highlights the obvious point that individuals are different. Why is it that some apparently have the ability to eat prodigious quantities of food without becoming obese whereas others do not? Do individual differences result in altered metabolic responses to diet; i.e., do variations in personal metabolic set points contribute to obesity? Two lines of evidence suggest that the prenatal environment has a role in establishing such set point differences. First, babies born to mothers who smoked while pregnant exhibited low birth weight and had an increased risk of obesity and metabolic syndrome later in life (Power & Jefferis, 2002; Oken *et al.*, 2005; Al Mamun *et al.*, 2006). Second, babies who received inadequate nutrition in utero grew up to become adults with higher rates of cardiovascular disease (Barker & Osmond, 1986, 1988; Barker *et al.*, 1989). The Developmental Origins of Health and Disease (DOHaD) model proposes that the prenatal and early life environment plays a key role in establishing life-long patterns of health and disease (Gluckman & Hanson, 2004). The DOHaD model holds that the development of chronic diseases (or the lack of chronic disease) is influenced by environmental factors (amount and quality of diet, chemical exposure, maternal stress, etc.) acting in early life that interact with genetic differences and factors associated with adult lifestyle. Simplistically, the prenatal environment elicits corresponding changes in the foetus that adjust metabolism to match the projected caloric environment. However, metabolic changes that favour the most effective use of scarce calories will not be adaptive when calories are in excess as in modern societies (Gluckman *et al.*, 2008). There is some evidence to suggest that 'catch-up growth' in early life is a key factor predisposing individuals towards obesity, insulin resistance and cardiovascular disease (Ong *et al.*, 2000; Ong & Dunger, 2002). Thus, while individuals appear outwardly normal, early life events occurring during critical developmental time windows (e.g., perinatally) may lead to permanent changes that are manifested at adulthood (Hanson & Gluckman, 2008). Although the bulk of evidence related to DOHaD comes from nutritional studies, there is no reason to suppose that other factors, such as prenatal stress (Entringer *et al.*, 2008, 2009) and exposure to endocrine disrupting chemicals (EDCs) (Janesick & Blumberg, 2011a), will not elicit similar changes in metabolic programming. Adipogenesis, weight control and metabolism are under hormonal control and are thus susceptible to interference by EDCs. EDC exposure has been linked with diabetes and metabolic syndrome, which may be related to, or independent from their effects on obesity (Lee *et al.*, 2010; Sergeev & Carpenter, 2010, 2011).

Obesogens

'Obesogens' are chemical compounds that can promote obesity by increasing the number of fat cells (and fat storage into existing fat cells), by changing the amount of calories burned at rest, by altering energy balance to favour storage of calories and by altering the mechanisms through which the body regulates appetite and satiety. Our environmental obesogen hypothesis proposes that a subset of EDCs could promote the development of obesity. Although initially controversial, the obesogen hypothesis has gained momentum in recent years with the identification of obesogenic chemicals that promote adipogenesis and obesity in animals and humans (Newbold *et al.*, 2009; Janesick & Blumberg, 2011a,b,c; La Merrill & Birnbaum, 2011; Tang-Peronard *et al.*, 2011). Perhaps most significantly, several classes of pharmaceutical drugs have been linked with weight gain and obesity in humans. Among these are thiazolidinedione anti-diabetic drugs (Larsen *et al.*, 2003; Rubenstrunk *et al.*, 2007), tricyclic antidepressants (Berken *et al.*, 1984), selective serotonin reuptake inhibitors (Fava, 2000) and atypical anti-psychotic drugs, such as olanzapine (Nemeroff, 1997). Considering that exposure to these drugs has been linked with obesity in humans, it is reasonable to suppose that exposure to EDCs targeting the same pathways will produce similar outcomes. For example, thiazolidinediones activate the peroxisome proliferator-activated receptor gamma (PPAR γ), a ligand-dependent transcription factor that is a key regulator of adipogenesis (Evans *et al.*, 2004; Tontonoz & Spiegelman, 2008). Chemicals that activate PPAR γ should have the same effect.

Indeed, we and others identified the organotin tributyltin (TBT) as a xenobiotic obesogen (Kanayama *et al.*, 2005; Grun *et al.*, 2006). TBT and the related chemical, triphenyltin (TPT), are nanomolar affinity ligands for PPAR γ and its heterodimeric partner, the retinoid X receptor (RXR) that were shown to induce adipogenesis in preadipocyte cell lines, such as 3T3-L1 cells (Kanayama *et al.*, 2005; Grun *et al.*, 2006). Currently, organotins are prevalent used in industry, as fungicides, wood preservatives and heat stabilizers in polyolefin plastics (Piver, 1973; Nath, 2008). Organotins, including TBT have been documented in house dust, suggesting that exposure from sources other than food may be widespread (Kannan *et al.*, 2010). Although TBT has largely been phased out of agricultural use, TPT remains in use as a fungicide and miticide. Organotins are lipophilic and have been shown to bioaccumulate in bacteria, algae and aquatic invertebrates (Hoch, 2001). Although TBT is most famous for its sex altering effects on gastropod mollusks (Blaber, 1970; Gibbs & Bryan, 1986) and fish (Shimasaki *et al.*, 2003), we unexpectedly found that *Xenopus laevis*

tadpoles exposed to low levels of TBT exhibited ectopic fat cell production (Grun *et al.*, 2006). In mice, prenatal exposure to TBT during gestation resulted in premature accumulation of fat in adipose tissues at birth and increased fat depot size at 10 weeks of age, although, the exposed mice were slightly smaller (Grun *et al.*, 2006). The main conclusion from these studies was that the tendency to store excess fat was programmed before birth due to TBT exposure. Subsequent experiments aimed at understanding the mechanisms underlying the effects of prenatal TBT exposure revealed that a single prenatal treatment with TBT or with the pharmaceutical obesogen, rosiglitazone (ROSI), altered the fate of multipotent mesenchymal stromal stem cells (MSCs). MSCs normally give rise to several tissue types *in vivo*, including bone, adipose and cartilage (Pittenger *et al.*, 1999). In offspring of pregnant dams treated with a single dose of TBT or ROSI, MSCs derived from white adipose tissue were predisposed to become adipocytes. MSCs derived from obesogen treated animals were about twice as likely to differentiate into adipocytes in culture as control cells and this frequency was further increased by subsequent *in vitro* treatment with TBT or ROSI (Kirchner *et al.*, 2010). The ability of these cells to differentiate into bone was correspondingly inhibited (Kirchner *et al.*, 2010). The ability of TBT or ROSI to induce adipogenesis in MSCs (Kirchner *et al.*, 2010) or in 3T3-L1 preadipocytes (Li *et al.*, 2011) was completely dependent on activation of PPAR γ , suggesting that the *in vivo* effects of these compounds similarly depend on PPAR γ . However, this remains to be demonstrated.

The topic of obesogens and obesogen action has been extensively reviewed in recent years (Grun & Blumberg, 2009a,b; Grun, 2010; Newbold, 2010, 2011; Blumberg, 2011; Heindel, 2011; Janesick & Blumberg, 2011a,b,c; Tang-Peronard *et al.*, 2011) as have the effects of EDCs on metabolism (Diamanti-Kandarakis *et al.*, 2009; Casals-Casas & Desvergne, 2011). In this review, we highlight likely mechanisms for obesogen action and summarize recent studies linking EDC exposure with obesity in humans.

Obesogens acting on sex steroid receptors

Estrogens in the adult are protective against abdominal obesity and metabolic disease whereas perinatal oestrogen exposure has the opposite effect (see below). Ovariectomized rats (a model for menopause in women) developed abdominal obesity, which was reversed upon treatment with oestrogen (Laudenslager *et al.*, 1980; Wade *et al.*, 1985). Consistent with this observation, loss-of-function in the oestrogen receptor alpha (ER α) resulted in increased white adipose depot size, central weight gain

and impaired glucose metabolism (Heine *et al.*, 2000; Cooke *et al.*, 2001). Knockout of P450 aromatase in mice inhibited the conversion of testosterone to estradiol, producing obese animals (Jones *et al.*, 2000); loss of the human CYP19A1 (aromatase) gene produced metabolic disease, fatty liver and abdominal obesity (Maffei *et al.*, 2007).

In contrast to its effects in adults, perinatal exposure to excess oestrogen promoted weight gain. Mice treated neonatally with the potent synthetic oestrogen diethylstilbesterol (DES) gave birth to pups that were initially smaller, but became heavier later in life (Newbold *et al.*, 2005, 2008, 2009; Newbold, 2010, 2011). Similarly, treatment of pregnant mouse (Cagampang *et al.*, 2007) or rat (Rubin *et al.*, 2001) dams with the environmental oestrogen bisphenol-A (BPA) resulted in smaller offspring that exhibited 'catch-up' growth and were significantly heavier as adults. Dichlorodiphenyl-dichloroethylene (DDE), the major metabolite of the pesticide DDT, is both an oestrogen receptor activator and an anti-androgen (Kupfer & Bulger, 1976; Kelce *et al.*, 1995). Mothers who lived along the Lake Michigan shoreline where they were exposed to high levels of DDT, were more likely to have a child that exhibited elevated BMI in adulthood (Karmaus *et al.*, 2009). More recently, Mendez and colleagues showed that prenatal exposure to DDE was associated with rapid weight gain in human infants and elevated BMI later in infancy (Mendez *et al.*, 2011). Despite a large number of available studies, the effects of BPA on health remain controversial. Recent human studies have revealed a link between BPA levels and obesity in humans (Carwile & Michels, 2011) and animal studies showed low dose effects of BPA on obesity and diabetes (Rubin, 2011). A recent study tested the effects of prenatal BPA exposure and concluded that while the animals were larger and males had significantly more fat stored at 7 weeks, the animals were neither obese nor did they have increased susceptibility to the effects of high fat diet at adulthood (Ryan *et al.*, 2010). The prevailing view at the moment is that low dose gestational BPA exposure is likely to be causally linked with the development of obesity. Although it appears likely that BPA exerts its obesogenic effects by acting as a developmental oestrogen, the mechanism(s) through which BPA acts to exert its deleterious effects on health need to be more fully elucidated. Ongoing studies in a number of laboratories should shed further light onto this important issue in the near future.

Obesogens and glucocorticoid metabolism

In addition to the sex steroid receptors, disruption of another nuclear hormone receptor regulated signalling pathway, the glucocorticoid receptor, is known to

contribute to obesity. Obesity is linked to a general increase of positive feedback within the hypothalamic-pituitary-adrenocortical (HPA) axis, leading to an over-secretion of cortisol from the adrenal gland (Marin *et al.*, 1992; Björntorp, 1993; Chalew *et al.*, 1995; Björntorp, 1997; Björntorp & Rosmond, 2000). However, rather than causing higher circulating glucocorticoid levels, obesity-related hypercortisolism is generally peripheral, local and characterized by an impaired ability to clear cortisol in adipose tissue, especially visceral adipose tissue (Rask *et al.*, 2001). Glucocorticoids increased both the differentiation of adipocytes from MSCs and the proliferation of adipocytes. Therefore, excess glucocorticoid levels in adipose depots are likely to stimulate local adipogenesis (Hauner *et al.*, 1989; Björntorp, 1991; Bujalska *et al.*, 1999).

One possible mechanism underlying peripheral hypercortisolism is dysregulation of 11 β -hydroxysteroid dehydrogenase type-1 (11 β HSD1), a ubiquitously expressed enzyme that primarily functions to convert inactive glucocorticoids, such as cortisone (humans) and 11-dehydrocorticosterone (rodents) into their active relatives cortisol and corticosterone (Seckl *et al.*, 2004). Elevated 11 β HSD1 has been linked with obesity and metabolic syndrome in humans (Rask *et al.*, 2001; Wake *et al.*, 2003; Valsamakis *et al.*, 2004) and in obese Zucker rats, (Livingstone *et al.*, 2000). Excess glucocorticoid exposure during pregnancy was often associated with lower birth weights, but increased risk of cardiovascular disease, diabetes and hypertension in the adult offspring (Seckl, 2001). Maternal stress has been linked with increased levels of corticotropin-releasing hormone, increased cortisol secretion and reduced birth weight in the offspring (Weinstock, 2005; Entringer *et al.*, 2009, 2010). Monkeys treated with the synthetic glucocorticoid dexamethasone during pregnancy produced offspring that were normal at birth, but exhibited significant weight gain at 2 months of age, subsequently became obese and developed metabolic syndrome (increased blood pressure, high total cholesterol, decreased HDL and insulin resistance) (Schlumbohm *et al.*, 2007).

Activity of the hypothalamic-pituitary-adrenocortical (HPA) axis that regulates glucocorticoid homeostasis is tightly regulated; therefore, it is possible that any EDC that perturbs the set point of this axis in early life could contribute to subsequent obesity. Such a mechanism could account, at least in part, for why many people cannot lose weight effectively. There are many possible mechanisms through which EDCs could modulate glucocorticoid homeostasis to disrupt energy balance, appetite and the stress response (Odermatt & Gumy, 2008). As 11 β HSD1 catalyzes the conversion of inactive to active glucocorticoids, increasing the activity of 11 β HSD1 could

readily disrupt the HPA axis. This is generally prevented in the foetus because placental 11 β HSD2, which catalyzes the conversion of active to inactive glucocorticoids, is highly expressed throughout pregnancy to reduce foetal cortisol exposure (Edwards *et al.*, 1993). Prenatal inhibition of 11 β HSD2 by carbenoxolone administration resulted in reduced birth weight, increased anxious behaviour and increased secretion of corticotropin-releasing hormone in rats (Welberg *et al.*, 2000). Therefore, increased glucocorticoid transport to the foetus by hyperactivating 11 β HSD1, or inhibiting 11 β HSD2 in the placenta are potential mechanisms through which EDCs might disrupt the HPA axis. Dithiocarbamates decreased 11 β HSD2 activity in vitro (Atanasov *et al.*, 2003) as did organotins (Atanasov *et al.*, 2005). Moreover, dibutyltin inhibited the binding of ligands to the glucocorticoid receptor and the ability of this receptor to inhibit cytokine activity and inflammation (Gumy *et al.*, 2008). Another potential mechanism for altered glucocorticoid homeostasis could be alterations in the levels of corticosteroid-binding globulin (CBG) (Fernandez-Real *et al.*, 2002). Adipose tissue that is deficient in CBG cannot evacuate excess cortisol to the blood; moreover, CBG deficiency in humans leads to increased proliferation and differentiation of pre-adipocytes into adipocytes (Joyner *et al.*, 2003). Therefore, exposure to EDCs that decrease CBG activity might also lead to obesity in the adult. Disruption of glucocorticoid action, stress and obesity are fertile areas for future studies because very little research has addressed EDCs and the HPA axis.

Peroxisome proliferator-activated receptors as obesogen targets

The peroxisome proliferator activated receptors (PPARs) are a family of nuclear hormone receptors that respond to fatty acids and related ligands (Casals-Casas *et al.*, 2008; Casals-Casas & Desvergne, 2011). There are three PPARs, PPAR α , PPAR β/δ and PPAR γ that all form obligate heterodimers with RXR to regulate the expression of target genes at the transcriptional level (Tontonoz & Spiegelman, 2008). PPAR γ is considered to be the master regulator of adipogenesis (Evans *et al.*, 2004) and plays key roles in nearly all aspects of adipocyte biology (Tontonoz & Spiegelman, 2008). Thiazolidinediones, which combat type 2 diabetes, are potent activators of PPAR γ (Lehmann *et al.*, 1995) and stimulation of PPAR γ -regulated transcription is obesogenic, per se (Janesick & Blumberg, 2011b). Therefore, PPAR γ has become a focus of many recent obesity-related studies. The ligand-binding pocket of PPAR γ is large (Nolte *et al.*, 1998) and can accommodate various chemical structures (Maloney & Waxman, 1999). The mechanistic basis for TBT-promoted

adipogenesis was most strongly supported by evidence that TBT is an agonist for both PPAR γ and RXR (Kanayama *et al.*, 2005; Grun *et al.*, 2006). Competitive binding assays showed that TBT has comparable affinity to synthetic RXR agonists for RXR (Grun *et al.*, 2006). The crystal structure of TBT along with the RXR α ligand binding domain, plus a coactivator fragment, showed that TBT binds covalently to RXR (le Maire *et al.*, 2009), which means that it will not readily dissociate once attached. It has been proposed that TBT acts through RXR to promote adipogenesis and obesity (le Maire *et al.*, 2009). However, treatment with the potent PPAR γ antagonist T0070907 (Lee *et al.*, 2002), inhibited TBT- or ROSI-stimulated adipogenesis in mouse and human MSCs (Kirchner *et al.*, 2010), whereas treatment with the related PPAR γ antagonist GW9662 blocked adipogenesis in 3T3-L1 preadipocytes (Li *et al.*, 2011). The conclusion from these studies was that the stimulation of adipogenesis in MSCs and cell lines by ROSI or TBT required activation of PPAR γ .

Because PPAR γ is the master regulator of adipogenesis, it is clear that activation of PPAR γ by EDCs is a potential risk factor for obesity. However, TBT is not the most common EDC to which humans are exposed. Phthalates are ubiquitous organic chemicals that give plastics, such as polyvinyl chloride (PVC), more flexibility and durability and readily leach into food, from medical devices and materials used in construction and manufacturing. Some phthalates were shown to be PPAR γ agonists (Hurst & Waxman, 2003) and stimulated the proliferation of adipocytes in the 3T3-L1 cell culture model (Feige *et al.*, 2007). Phthalate metabolites were associated with increased waist circumference in men (Stahlhut *et al.*, 2007), and therefore, are predicted to be obesogenic. It is quite likely that other xenobiotic chemicals activate PPAR γ and may contribute to the aetiology of obesity (Janesick & Blumberg, 2011b). Screening efforts such as the EPA's Toxcast (Dix *et al.*, 2007; Knudsen *et al.*, 2011) and the joint NIEHS/EPA/FDA Tox21 (Shukla *et al.*, 2010) are likely sources for the identification of new obesogens that act on PPAR γ and other biological targets.

Intriguingly, it has recently been shown that in addition to its known effects in adipocytes and MSCs, PPAR γ plays an important role in the brain by controlling appetite and metabolism in response to a high fat diet (Lu *et al.*, 2011; Myers & Burant, 2011; Ryan *et al.*, 2011). Specifically activating PPAR γ in the brain lead to increased feeding and accrued body weight whereas blockade of PPAR γ or PPAR γ loss-of-function lead to decreased consumption of high fat, but not normal diet. The conclusion from these studies was that tissue-specific regulation of PPAR γ action may play an important role in the outcome of exposure to chemicals that regulate

PPAR γ and in the body's response to dietary excesses. The identification of other PPAR γ disruptors, as well as the molecular pathways targeted by EDC-PPAR γ action that reprogram stem cell fate to favour obesity will be important areas for future research.

Epigenetics connects environmental exposures with gene expression

Obesogens are predicted to act prenatally by eliciting epigenetic modifications that alter the expression of key genes in adipogenic pathways. Epigenetic modifications in genes responsible for regulating metabolism, body weight and fat deposition could result in developmental plasticity that allow an organism to make rapid adaptations to changing environments, typically by altering levels of gene expression via DNA methylation or modification of histone proteins (Gluckman & Hanson, 2004; Godfrey *et al.*, 2007, 2011; Gluckman *et al.*, 2008; Hanson & Gluckman, 2008; Hanson *et al.*, 2011). Epigenetic changes that occur during germ cell development can potentially lead to transgenerational effects that may persist for many generations after the initial exposure (Skinner, 2010; Skinner *et al.*, 2011).

Numerous studies have shown that changes in the nutritional environment lead to alterations in the methylation status of genes (Burdge & Lillycrop, 2010a,b; Jackson *et al.*, 2010; Lillycrop & Burdge, 2011; Godfrey *et al.*, 2011; Hochberg *et al.*, 2011; Lillycrop, 2011). Foetal liver derived from rats fed a low-protein diet showed promoter hypermethylation in the liver X-receptor (LXR) (van Straten *et al.*, 2010) and hypomethylation of PPAR α (Lillycrop *et al.*, 2008). The methylation-deficient status of PPAR α was rescued by supplementing the low-protein diet with the methyl donor, folic acid (Lillycrop *et al.*, 2008). Increased methylation of the RXR α promoter in humans was associated with increased fat mass at 9 years of age (Godfrey *et al.*, 2011). Considered together, it is reasonable to infer that such epigenetic changes could lead to disturbances in metabolism and lipid homeostasis that might be causally linked to obesity. Further studies will be illuminating in this regard.

If changes in prenatal nutrition can lead to epigenetic changes, does exposure to EDCs elicit the same effects? Consistent with this possibility, chemical-induced alterations in DNA methylation status were observed for diethylstilbestrol (DES) (Li *et al.*, 1997), TCDD (Wu *et al.*, 2004), vinclozolin (Anway *et al.*, 2005), BPA (Bernal & Jirtle, 2010) and TBT (Kirchner *et al.*, 2010). DNA methyltransferase activity was altered in rat embryos, depending on whether the embryo was exposed to TCDD, DES or polychlorinated biphenyl-153 (PCB153) (Wu *et al.*, 2006). Thus, although the evidence is still emerging, EDCs can affect the expression levels of

DNA and histone methyltransferases that might lead to subsequent broad impacts on gene expression, including genes that are important for metabolism and obesity.

Although the potential for EDCs to alter epigenetic programming to favour altered gene expression definitely exists (Jackson *et al.*, 2010; Lillycrop & Burdge, 2011; Hochberg *et al.*, 2011; Lillycrop, 2011) evidence supporting specific mechanisms of action is scant. One mechanism that has been described concerns epigenetic modifications that alter stem cell fate (Kirchner *et al.*, 2010; Janesick & Blumberg, 2011a). Adipogenesis is a differentiation event in the mesodermal lineage in which MSCs or more lineage-restricted derivatives give rise to adipocytes. MSCs harvested from epididymal or ovarian fat pads of mice exposed to TBT in utero differentiated into significantly more fat cells, compared with controls, whereas fewer MSCs could differentiate into osteocytes (Kirchner *et al.*, 2010). TBT likely induced epigenetic changes within the MSC compartment that promoted demethylation of adipogenic genes, thereby biasing the MSC compartment to favour the adipocyte lineage (Kirchner *et al.*, 2010). Uninduced MSCs harvested from mice exposed to TBT in utero showed decreased methylation in the gene encoding fatty acid binding protein 4 (FABP4), a marker of adipocytes; suggesting that the MSC population had already been epigenetically modified to favour adipogenesis (Kirchner *et al.*, 2010). Future studies will be needed to identify which regulatory genes have had their expression altered by prenatal exposure to TBT and other obesogens, whether these are the result of epigenetic changes and if the changes elicited persist in future generations.

Conclusions and future prospects

Unhealthy food consumed in excessive amounts and insufficient physical activity are undoubtedly associated with obesity. Whether these are the major and proximate causes of obesity, as is commonly believed, or whether there are other significant causes for obesity remain to be demonstrated. Moreover, it currently remains unknown to what extent obesogen exposure interacts with dietary excesses and lifestyle factors to affect obesity. It is indisputable that following commonly espoused nutritional guidelines (decreased fat consumption, increased carbohydrate consumption) has not resulted in a leaner population. Rather, the opposite is true; we now have an epidemic of obesity in infants (Kim *et al.*, 2006), as well as in children and adults. This suggests that obesity is being programmed prenatally or in early childhood. There is increasing evidence that supports the proposal that environmental endocrine disrupting chemicals (Janesick & Blumberg, 2011a), together with calorie-dense

modern diets (Lustig, 2006) may contribute to the early life programming of obesity. Prenatal exposure to obesogens is likely to be an underestimated contributor to the obesity epidemic; moreover, a variety of persistent organic pollutants have been linked with obesity in human studies (Carwile & Michels, 2011; Lee *et al.*, 2011a,b, 2012; Mendez *et al.*, 2011; Tang-Peronard *et al.*, 2011). It will be important in the future to determine which of these chemicals are causally linked with adipogenesis and obesity using studies in appropriate animal models. Prenatal exposure to TBT, a chemical for which the mechanism of action is known, predisposed exposed individuals to produce more fat cells (Kirchner *et al.*, 2010) and accrue increased adipose depot mass (Grun *et al.*, 2006). This suggests that the DOHaD model is applicable to the effects of chemical exposure.

There are numerous EDCs (e.g., BPA, brominated flame retardants and phthalates), more prevalent in the environment than TBT that have been linked to metabolic disease (Casals-Casas *et al.*, 2008; Desvergne *et al.*, 2009; Rubin & Soto, 2009; Vandenberg *et al.*, 2009; Eskenazi *et al.*, 2011; Harley *et al.*, 2011; Rubin, 2011). The metabolic pathways targeted by most of these chemicals remain to be determined; although, some likely pathways are currently under study. The establishment of firm links between EDC exposure and obesity will require elucidation of the underlying mechanisms. Understanding how chemicals enter the body and are transferred to the developing foetus is still not well understood and requires further study. Determining the epigenetic basis of how early life exposure to EDCs modulates the developmental programming of future health and disease will provide answers to the mechanistic questions regarding how obesogens disrupt the endocrine system. There is much yet to learn about how EDC exposures reprogram stem cell fate to favour obesity and diabetes and to what extent these effects can be reduced or eliminated by dietary, behavioural or pharmaceutical interventions.

Acknowledgements

Work in the authors' laboratory was supported by a grant from the NIH R01 ES015849. A.J. is a pre-doctoral trainee of NSF IGERT DGE 0549479.

References

- Al Mamun A, Lawlor DA, Alati R, O'Callaghan MJ, Williams GM & Najman JM. (2006) Does maternal smoking during pregnancy have a direct effect on future offspring obesity? Evidence from a prospective birth cohort study. *Am J Epidemiol* 164, 317–325.
- Anway MD, Cupp AS, Uzumcu M & Skinner MK. (2005) Epigenetic transgenerational actions of endocrine disruptors and male fertility. *Science* 308, 1466–1469.

- Atanasov AG, Tam S, Rocken JM, Baker ME & Odermatt A. (2003) Inhibition of 11 beta-hydroxysteroid dehydrogenase type 2 by dithiocarbamates. *Biochem Biophys Res Commun* 308, 257–262.
- Atanasov AG, Nashev LG, Tam S, Baker ME & Odermatt A. (2005) Organotins disrupt the 11beta-hydroxysteroid dehydrogenase type 2-dependent local inactivation of glucocorticoids. *Environ Health Perspect* 113, 1600–1606.
- Barker DJ & Osmond C. (1986) Infant mortality, childhood nutrition, and ischaemic heart disease in England and Wales. *Lancet* 1, 1077–1081.
- Barker DJ & Osmond C. (1988) Low birth weight and hypertension. *BMJ* 297, 134–135.
- Barker DJ, Osmond C, Golding J, Kuh D & Wadsworth ME. (1989) Growth *in utero*, blood pressure in childhood and adult life, and mortality from cardiovascular disease. *BMJ* 298, 564–567.
- Berken GH, Weinstein DO & Stern WC. (1984) Weight gain. A side-effect of tricyclic antidepressants. *J Affect Disord* 7, 133–138.
- Bernal AJ & Jirtle RL. (2010) Epigenomic disruption: the effects of early developmental exposures. *Birth Defects Res A Clin Mol Teratol* 88, 938–944.
- Bjorntorp P. (1991) Adipose tissue distribution and function. *Int J Obes* 15(Suppl. 2), 67–81.
- Bjorntorp P. (1997) Neuroendocrine factors in obesity. *J Endocrinol* 155, 193–195.
- Björntorp P. (1993) Visceral obesity: a “Civilization syndrome”. *Obes Res* 1, 206–222.
- Bjorntorp P & Rosmond R. (2000) Obesity and cortisol. *Nutrition* 16, 924–936.
- Blaber SJM. (1970) The occurrence of a penis-like outgrowth behind the right tentacle in spent females of *Nucella lapillus*. *Proc Malacolog Soc Lon* 39, 231–233.
- Blumberg B. (2011) Obesogens, stem cells and the maternal programming of obesity. *J Develop Orig Health Dis* 2, 3–8.
- Bujalska IJ, Kumar S, Hewison M & Stewart PM. (1999) Differentiation of adipose stromal cells: the roles of glucocorticoids and 11beta-hydroxysteroid dehydrogenase. *Endocrinology* 140, 3188–3196.
- Burdge GC & Lillycrop KA. (2010a) Bridging the gap between epigenetics research and nutritional public health interventions. *Genome Med* 2, 80.
- Burdge GC & Lillycrop KA. (2010b) Nutrition, epigenetics, and developmental plasticity: implications for understanding human disease. *Annu Rev Nutr* 30, 315–339.
- Butte NF, Christiansen E & Sorensen TI. (2007) Energy imbalance underlying the development of childhood obesity. *Obesity (Silver Spring)* 15, 3056–3066.
- Cagampan F, Anthony F & Hanson M. (2007) P2-14 developmental exposure to bisphenol-a leads to obesity and cardiovascular dysfunction in adult mouse offspring. *Early Hum Dev* 83, S132–S133.
- Carwile JL & Michels KB. (2011) Urinary bisphenol a and obesity: NHANES 2003–2006. *Environ Res* 111, 825–830.
- Casals-Casas C & Desvergne B. (2011) Endocrine disruptors: from endocrine to metabolic disruption. *Annu Rev Physiol* 73, 135–162.
- Casals-Casas C, Feige JN & Desvergne B. (2008) Interference of pollutants with PPARs: endocrine disruption meets metabolism. *Int J Obes (Lond)* 32(Suppl. 6), S53–S61.
- Chalew S, Nagel H & Shore S. (1995) The hypothalamic-pituitary-adrenal axis in obesity. *Obes Res* 3, 371–382.
- Cooke PS, Heine PA, Taylor JA & Lubahn DB. (2001) The role of estrogen and estrogen receptor- α in male adipose tissue. *Mol Cell Endocrinol* 178, 147–154.
- Desvergne B, Feige JN & Casals-Casas C. (2009) PPAR-mediated activity of phthalates: a link to the obesity epidemic? *Mol Cell Endocrinol* 304, 43–48.
- Diamanti-Kandarakis E, Bourguignon JP, Giudice LC, Hauser R, Prins GS, Soto AM, Zoeller RT & Gore AC. (2009) Endocrine-disrupting chemicals: an endocrine society scientific statement. *Endocr Rev* 30, 293–342.
- Dix DJ, Houck KA, Martin MT, Richard AM, Setzer RW & Kavlock RJ. (2007) The ToxCast program for prioritizing toxicity testing of environmental chemicals. *Toxicol Sci* 95, 5–12.
- Edwards CR, Benediktsson R, Lindsay RS & Seckl JR. (1993) Dysfunction of placental glucocorticoid barrier: link between fetal environment and adult hypertension? *Lancet* 341, 355–357.
- Entringer S, Wust S, Kumsta R, Layes IM, Nelson EL, Hellhammer DH & Wadhwa PD. (2008) Prenatal psychosocial stress exposure is associated with insulin resistance in young adults. *Am J Obstet Gynecol* 199, 498 e491–497.
- Entringer S, Kumsta R, Hellhammer DH, Wadhwa PD & Wust S. (2009) Prenatal exposure to maternal psychosocial stress and HPA axis regulation in young adults. *Horm Behav* 55, 292–298.
- Entringer S, Buss C & Wadhwa PD. (2010) Prenatal stress and developmental programming of human health and disease risk: concepts and integration of empirical findings. *Curr Opin Endocrinol Diabetes Obes* 17, 507–516.
- Eskenazi B, Fenster L, Castorina R, Marks AR, Sjodin A, Rosas LG, Holland N, Guerra AG, Lopez-Carrillo L & Bradman A. (2011) A comparison of PBDE serum concentrations in Mexican and Mexican-American children living in California. *Environ Health Perspect* 119, 1442–1448.
- Evans RM, Barish GD & Wang YX. (2004) PPARs and the complex journey to obesity. *Nat Med* 10, 355–361.
- Fava M. (2000) Weight gain and antidepressants. *J Clin Psychiatry* 61(Suppl. 11), 37–41.
- Feige JN, Gelman L, Rossi D, Zoete V, Metivier R, Tudor C *et al.* (2007) The endocrine disruptor monoethyl-hexyl-phthalate is a selective peroxisome proliferator-activated receptor gamma modulator that promotes adipogenesis. *J Biol Chem* 282, 19152–19166.
- Fernandez-Real JM, Pugeat M, Grasa M, Broch M, Vendrell J, Brun J & Ricart W. (2002) Serum corticosteroid-binding globulin concentration and insulin resistance syndrome: a population study. *J Clin Endocrinol Metab* 87, 4686–4690.
- Flegal KM, Carroll MD, Ogden CL & Curtin LR. (2010) Prevalence and trends in obesity among US adults, 1999–2008. *JAMA* 303, 235–241.
- Freedland ES. (2004) Role of a critical visceral adipose tissue threshold (CVAT) in metabolic syndrome: implications for controlling dietary carbohydrates: a review. *Nutr Metab (Lond)* 1, 12.
- Gibbs P & Bryan G. (1986) Reproductive failure in populations of the dog-whelk, *Nucella lapillus*, caused by imposex induced by tributyltin from antifouling paints. *J Mar Biol Assoc UK* 66, 767–777.
- Gluckman PD & Hanson MA. (2004) Living with the past: evolution, development, and patterns of disease. *Science* 305, 1733–1736.
- Gluckman PD, Hanson MA, Beedle AS & Raubenheimer D. (2008) Fetal and neonatal pathways to obesity. *Front Horm Res* 36, 61–72.
- Godfrey KM, Lillycrop KA, Burdge GC, Gluckman PD & Hanson MA. (2007) Epigenetic mechanisms and the mismatch concept of the developmental origins of health and disease. *Pediatr Res* 61, 5R–10R.
- Godfrey KM, Sheppard A, Gluckman PD, Lillycrop KA, Burdge GC, McLean C *et al.* (2011) Epigenetic gene promoter methylation at

- birth is associated with child's later adiposity. *Diabetes* 60, 1528–1534.
- Grun F. (2010) Obesogens. *Curr Opin Endocrinol Diabetes Obes* 17, 453–459.
- Grun F & Blumberg B. (2009a) Endocrine disrupters as obesogens. *Mol Cell Endocrinol* 304, 19–29.
- Grun F & Blumberg B. (2009b) Minireview: the case for obesogens. *Mol Endocrinol* 23, 1127–1134.
- Grun F, Watanabe H, Zamanian Z, Maeda L, Arima K, Cubacha R, Gardiner DM, Kanno J, Iguchi T & Blumberg B. (2006) Endocrine-disrupting organotin compounds are potent inducers of adipogenesis in vertebrates. *Mol Endocrinol* 20, 2141–2155.
- Gumy C, Chandsawangbhuwana C, Dzykanchuk AA, Kratschmar DV, Baker ME & Odermatt A. (2008) Dibutyltin disrupts glucocorticoid receptor function and impairs glucocorticoid-induced suppression of cytokine production. *PLoS ONE* 3, e3545.
- Hanson MA & Gluckman PD. (2008) Developmental origins of health and disease: new insights. *Basic Clin Pharmacol Toxicol* 102, 90–93.
- Hanson M, Godfrey KM, Lillycrop KA, Burdge GC & Gluckman PD. (2011) Developmental plasticity and developmental origins of non-communicable disease: theoretical considerations and epigenetic mechanisms. *Prog Biophys Mol Biol* 106, 272–280.
- Harley KG, Chevrier J, Schall RA, Sjodin A, Bradman A & Eskenazi B. (2011) Association of prenatal exposure to polybrominated diphenyl ethers and infant birth weight. *Am J Epidemiol* 174, 885–892.
- Hauner H, Entenmann G, Wabitsch M, Gaillard D, Ailhaud G, Negrel R & Pfeiffer EF. (1989) Promoting effect of glucocorticoids on the differentiation of human adipocyte precursor cells cultured in a chemically defined medium. *J Clin Invest* 84, 1663–1670.
- Heindel JJ. (2011) The obesogen hypothesis of obesity: overview and human evidence. In: *Obesity Before Birth*, Vol. 30. (ed. RH Lustig), pp. 355–366. Springer, US.
- Heine PA, Taylor JA, Iwamoto GA, Lubahn DB & Cooke PS. (2000) Increased adipose tissue in male and female estrogen receptor- α knockout mice. *Proc Natl Acad Sci U S A* 97, 12729–12734.
- Hoch M. (2001) Organotin compounds in the environment – an overview. *Appl Geochem* 16, 719–743.
- Hochberg Z, Feil R, Constanca M, Fraga M, Junien C, Carel JC *et al.* (2011) Child health, developmental plasticity, and epigenetic programming. *Endocr Rev* 32, 159–224.
- Hurst CH & Waxman DJ. (2003) Activation of ppar α and ppar γ by environmental phthalate monoesters. *Toxicol Sci* 74, 297–308.
- Ibrahim MM. (2010) Subcutaneous and visceral adipose tissue: structural and functional differences. *Obes Rev* 11, 11–18.
- Jackson AA, Burdge GC & Lillycrop KA. (2010) Diet, nutrition and modulation of genomic expression in fetal origins of adult disease. *World Rev Nutr Diet* 101, 56–72.
- Janesick A & Blumberg B. (2011a) Endocrine disrupting chemicals and the developmental programming of adipogenesis and obesity. *Birth Defects Res C Embryo Today* 93, 34–50.
- Janesick A & Blumberg B. (2011b) Minireview: Ppargamma as the target of obesogens. *J Steroid Biochem Mol Biol* 127, 4–8.
- Janesick A & Blumberg B. (2011c) The role of environmental obesogens in the obesity epidemic. In: *Obesity Before Birth*, Vol. 30. (ed. RH Lustig), pp. 383–399. Springer, US.
- Jones ME, Thorburn AW, Britt KL, Hewitt KN, Wreford NG, Proietto J *et al.* (2000) Aromatase-deficient (arko) mice have a phenotype of increased adiposity. *Proc Natl Acad Sci U S A* 97, 12735–12740.
- Joyner JM, Hutley LJ, Bachmann AW, Torpy DJ & Prins JB. (2003) Greater replication and differentiation of preadipocytes in inherited corticosteroid-binding globulin deficiency. *Am J Physiol Endocrinol Metab* 284, E1049–E1054.
- Kanayama T, Kobayashi N, Mamiya S, Nakanishi T & Nishikawa J. (2005) Organotin compounds promote adipocyte differentiation as agonists of the peroxisome proliferator-activated receptor gamma/retinoid x receptor pathway. *Mol Pharmacol* 67, 766–774.
- Kannan K, Takahashi S, Fujiwara N, Mizukawa H & Tanabe S. (2010) Organotin compounds, including butyltins and octyltins, in house dust from albany, New York, USA. *Arch Environ Contam Toxicol* 58, 901–907.
- Karmaus W, Osuch JR, Eneli I, Mudd LM, Zhang J, Mikucki D, Haan P & Davis S. (2009) Maternal levels of dichlorodiphenyl-dichloroethylene (dde) may increase weight and body mass index in adult female offspring. *Occup Environ Med* 66, 143–149.
- Kelce WR, Stone CR, Laws SC, Gray LE, Kempainen JA & Wilson EM. (1995) Persistent ddt metabolite p,p'-dde is a potent androgen receptor antagonist. *Nature* 375, 581–585.
- Kim HK, Nelson-Dooley C, Della-Fera MA, Yang JY, Zhang W, Duan J, Hartzell DL, Hamrick MW & Baile CA. (2006) Genistein decreases food intake, body weight, and fat pad weight and causes adipose tissue apoptosis in ovariectomized female mice. *J Nutr* 136, 409–414.
- Kirchner S, Kieu T, Chow C, Casey S & Blumberg B. (2010) Prenatal exposure to the environmental obesogen tributyltin predisposes multipotent stem cells to become adipocytes. *Mol Endocrinol* 24, 526–539.
- Knudsen TB, Houck KA, Sipes NS, Singh AV, Judson RS, Martin MT *et al.* (2011) Activity profiles of 309 toxic chemicals evaluated across 292 biochemical targets. *Toxicology* 282, 1–15.
- Kupfer D & Bulger WH. (1976) Studies on the mechanism of estrogenic actions of o,p'-ddt: interactions with the estrogen receptor. *Pesticide Biochem Physiol* 6, 561–570.
- La Merrill M & Birnbaum LS. (2011) Childhood obesity and environmental chemicals. *Mt Sinai J Med* 78, 22–48.
- Larsen TM, Toubro S & Astrup A. (2003) Ppargamma agonists in the treatment of type ii diabetes: is increased fatness commensurate with long-term efficacy? *Int J Obes Relat Metab Disord* 27, 147–161.
- Laudenslager ML, Wilkinson CW, Carlisle HJ & Hammel HT. (1980) Energy balance in ovariectomized rats with and without estrogen replacement. *Am J Physiol* 238, R400–R405.
- Lee G, Elwood F, McNally J, Weiszmann J, Lindstrom M, Amaral K *et al.* (2002) T0070907, a selective ligand for peroxisome proliferator-activated receptor gamma, functions as an antagonist of biochemical and cellular activities. *J Biol Chem* 277, 19649–19657.
- Lee DH, Steffes MW, Sjodin A, Jones RS, Needham LL & Jacobs DR Jr. (2010) Low dose of some persistent organic pollutants predicts type 2 diabetes: a nested case-control study. *Environ Health Perspect* 118, 1235–1242.
- Lee DH, Lind PM, Jacobs DR Jr, Salihovic S, van Bavel B & Lind L. (2011a) Polychlorinated biphenyls and organochlorine pesticides in plasma predict development of type 2 diabetes in the elderly: the prospective investigation of the vasculature in uppsala seniors (pivus) study. *Diabetes Care* 34, 1778–1784.
- Lee DH, Steffes MW, Sjodin A, Jones RS, Needham LL & Jacobs DR Jr. (2011b) Low dose organochlorine pesticides and polychlorinated biphenyls predict obesity, dyslipidemia, and insulin resistance among people free of diabetes. *PLoS ONE* 6, e15977.
- Lee DH, Lind L, Jacobs DR Jr, Salihovic S, van Bavel B & Lind PM. (2012) Associations of persistent organic pollutants with abdominal

- obesity in the elderly: the prospective investigation of the vasculature in uppsala seniors (pivus) study. *Environ Int* 40, 170–178.
- Lehmann JM, Moore LB, Smith-Oliver TA, Wilkison WO, Willson TM & Kliewer SA. (1995) An antidiabetic thiazolidinedione is a high affinity ligand for peroxisome proliferator-activated receptor gamma (ppar gamma). *J Biol Chem* 270, 12953–12956.
- Li S, Washburn KA, Moore R, Uno T, Teng C, Newbold RR, McLachlan JA & Negishi M. (1997) Developmental exposure to diethylstilbestrol elicits demethylation of estrogen-responsive lactoferrin gene in mouse uterus. *Cancer Res* 57, 4356–4359.
- Li X, Ycaza J & Blumberg B. (2011) The environmental obesogen tributyltin chloride acts via peroxisome proliferator activated receptor gamma to induce adipogenesis in murine 3T3-L1 preadipocytes. *J Steroid Biochem Mol Biol* 127, 9–15.
- Lillicrop KA. (2011) Effect of maternal diet on the epigenome: implications for human metabolic disease. *Proc Nutr Soc* 70, 64–72.
- Lillicrop KA & Burdge GC. (2011) Epigenetic changes in early life and future risk of obesity. *Int J Obes (Lond)* 35, 72–83.
- Lillicrop KA, Phillips ES, Torrens C, Hanson MA, Jackson AA & Burdge GC. (2008) Feeding pregnant rats a protein-restricted diet persistently alters the methylation of specific cytosines in the hepatic ppar alpha promoter of the offspring. *Br J Nutr* 100, 278–282.
- Livingstone DE, Jones GC, Smith K, Jamieson PM, Andrew R, Kenyon CJ & Walker BR. (2000) Understanding the role of glucocorticoids in obesity: tissue-specific alterations of corticosterone metabolism in obese Zucker rats. *Endocrinology* 141, 560–563.
- Lu M, Sarruff DA, Talukdar S, Sharma S, Li P, Bandyopadhyay G *et al.* (2011) Brain ppar-gamma promotes obesity and is required for the insulin-sensitizing effect of thiazolidinediones. *Nat Med* 17, 618–622.
- Lustig RH. (2006) Childhood obesity: behavioral aberration or biochemical drive? Reinterpreting the first law of thermodynamics. *Nat Clin Pract Endocrinol Metab* 2, 447–458.
- Lustig RH. (2010) Fructose: metabolic, hedonic, and societal parallels with ethanol. *J Am Diet Assoc* 110, 1307–1321.
- Maffei L, Rochira V, Zirilli L, Antunez P, Aranda C, Fabre B *et al.* (2007) A novel compound heterozygous mutation of the aromatase gene in an adult man: reinforced evidence on the relationship between congenital oestrogen deficiency, adiposity and the metabolic syndrome. *Clin Endocrinol (Oxf)* 67, 218–224.
- le Maire A, Grimaldi M, Roecklin D, Dagnino S, Vivat-Hannah V, Balaguer P & Bourguet W. (2009) Activation of rxr-ppar heterodimers by organotin environmental endocrine disruptors. *EMBO Rep* 10, 367–373.
- Maloney EK & Waxman DJ. (1999) Trans-activation of pparalpha and ppargamma by structurally diverse environmental chemicals. *Toxicol Appl Pharmacol* 161, 209–218.
- Marin P, Darin N, Amemiya T, Andersson B, Jern S & Bjorntorp P. (1992) Cortisol secretion in relation to body fat distribution in obese premenopausal women. *Metabolism* 41, 882–886.
- Mendez MA, Garcia-Esteban R, Guxens M, Vrijheid M, Kogevinas M, Goni F, Fochs S & Sunyer J. (2011) Prenatal organochlorine compound exposure, rapid weight gain, and overweight in infancy. *Environ Health Perspect* 119, 272–278.
- Muhlhauser B & Smith SR. (2009) Early-life origins of metabolic dysfunction: role of the adipocyte. *Trends Endocrinol Metab* 20, 51–57.
- Myers MG Jr & Burant CF. (2011) Ppar-gamma action: it's all in your head. *Nat Med* 17, 544–545.
- Nath M. (2008) Toxicity and the cardiovascular activity of organotin compounds: a review. *Appl Organomet Chem* 22, 598–612.
- Nemeroff CB. (1997) Dosing the antipsychotic medication olanzapine. *J Clin Psychiatry* 58(Suppl. 10), 45–49.
- Newbold RR. (2010) Impact of environmental endocrine disrupting chemicals on the development of obesity. *Hormones (Athens)* 9, 206–217.
- Newbold RR. (2011) Perinatal exposure to endocrine disrupting chemicals with estrogenic activity and the development of obesity. In: *Obesity Before Birth*, Vol. 30. (ed. RH Lustig), pp. 367–382. Springer, US.
- Newbold RR, Padilla-Banks E, Snyder RJ & Jefferson WN. (2005) Developmental exposure to estrogenic compounds and obesity. *Birth Defects Res A Clin Mol Teratol* 73, 478–480.
- Newbold RR, Padilla-Banks E, Jefferson WN & Heindel JJ. (2008) Effects of endocrine disruptors on obesity. *Int J Androl* 31, 201–208.
- Newbold RR, Padilla-Banks E & Jefferson WN. (2009) Environmental estrogens and obesity. *Mol Cell Endocrinol* 304, 84–89.
- Nolte RT, Wisely GB, Westin S, Cobb JE, Lambert MH, Kurokawa R, Rosenfeld MG, Willson TM, Glass CK & Milburn MV. (1998) Ligand binding and co-activator assembly of the peroxisome proliferator-activated receptor-gamma. *Nature* 395, 137–143.
- Odermatt A & Gummy C. (2008) Glucocorticoid and mineralocorticoid action: why should we consider influences by environmental chemicals? *Biochem Pharmacol* 76, 1184–1193.
- Oken E, Huh SY, Taveras EM, Rich-Edwards JW & Gillman MW. (2005) Associations of maternal prenatal smoking with child adiposity and blood pressure. *Obes Res* 13, 2021–2028.
- Ong KK & Dunger DB. (2002) Perinatal growth failure: the road to obesity, insulin resistance and cardiovascular disease in adults. *Best Pract Res Clin Endocrinol Metab* 16, 191–207.
- Ong KK, Ahmed ML, Emmett PM, Preece MA & Dunger DB. (2000) Association between postnatal catch-up growth and obesity in childhood: prospective cohort study. *BMJ* 320, 967–971.
- Pittenger MF, Mackay AM, Beck SC, Jaiswal RK, Douglas R, Mosca JD, Moorman MA, Simonetti DW, Craig S & Marshak DR. (1999) Multilineage potential of adult human mesenchymal stem cells. *Science* 284, 143–147.
- Piver WT. (1973) Organotin compounds: industrial applications and biological investigation. *Environ Health Perspect* 4, 61–79.
- Power C & Jefferis BJ. (2002) Fetal environment and subsequent obesity: a study of maternal smoking. *Int J Epidemiol* 31, 413–419.
- Rask E, Olsson T, Soderberg S, Andrew R, Livingstone DE, Johnson O & Walker BR. (2001) Tissue-specific dysregulation of cortisol metabolism in human obesity. *J Clin Endocrinol Metab* 86, 1418–1421.
- Rubenstrunk A, Hanf R, Hum DW, Fruchart JC & Staels B. (2007) Safety issues and prospects for future generations of ppar modulators. *Biochim Biophys Acta* 1771, 1065–1081.
- Rubin BS. (2011) Bisphenol a: an endocrine disruptor with widespread exposure and multiple effects. *J Steroid Biochem Mol Biol* 127, 27–34.
- Rubin BS & Soto AM. (2009) Bisphenol a: perinatal exposure and body weight. *Mol Cell Endocrinol* 304, 55–62.
- Rubin BS, Murray MK, Damassa DA, King JC & Soto AM. (2001) Perinatal exposure to low doses of bisphenol a affects body weight, patterns of estrous cyclicity, and plasma lh levels. *Environ Health Perspect* 109, 675–680.
- Ryan KK, Haller AM, Sorrell JE, Woods SC, Jandacek RJ & Seeley RJ. (2010) Perinatal exposure to bisphenol-a and the development of metabolic syndrome in cd-1 mice. *Endocrinology* 151, 2603–2612.
- Ryan KK, Li B, Grayson BE, Matter EK, Woods SC & Seeley RJ. (2011) A role for central nervous system ppar-gamma in the regulation of energy balance. *Nat Med* 17, 623–626.

- Schlumbohm C, Bramlage C, Strutz F, Armstrong VW, Oellerich M & Fuchs E. (2007) Predictive value of maternal bodyweight, postnatal weight gain and prenatal dexamethasone overexposure for the development of obesity in adult marmoset monkeys. *Exp Clin Endocrinol Diabetes* 115, 8.
- Seckl JR. (2001) Glucocorticoid programming of the fetus; adult phenotypes and molecular mechanisms. *Mol Cell Endocrinol* 185, 61–71.
- Seckl JR, Morton NM, Chapman KE & Walker BR. (2004) Glucocorticoids and 11beta-hydroxysteroid dehydrogenase in adipose tissue. *Recent Prog Horm Res* 59, 359–393.
- Sergeev AV & Carpenter DO. (2010) Increased hospitalizations for ischemic stroke with comorbid diabetes and residential proximity to sources of organic pollutants: a 12-year population-based study. *Neuroepidemiology* 35, 196–201.
- Sergeev AV & Carpenter DO. (2011) Increase in metabolic syndrome-related hospitalizations in relation to environmental sources of persistent organic pollutants. *Int J Environ Res Public Health* 8, 762–776.
- Shimasaki Y, Kitano T, Oshima Y, Inoue S, Imada N & Honjo T. (2003) Tributyltin causes masculinization in fish. *Environ Toxicol Chem* 22, 141–144.
- Shukla SJ, Huang R, Austin CP & Xia M. (2010) The future of toxicity testing: a focus on *in vitro* methods using a quantitative high-throughput screening platform. *Drug Discov Today* 15, 997–1007.
- Skinner MK. (2010) Metabolic disorders: fathers' nutritional legacy. *Nature* 467, 922–923.
- Skinner MK, Manikkam M & Guerrero-Bosagna C. (2011) Epigenetic transgenerational actions of endocrine disruptors. *Reprod Toxicol* 31, 337–343.
- Stahlhut RW, van Wijgaarden E, Dye TD, Cook S & Swan SH. (2007) Concentrations of urinary phthalate metabolites are associated with increased waist circumference and insulin resistance in adult U.S. Males. *Environ Health Perspect* 115, 876–882.
- van Straten EME, Bloks VW, Huijckman NCA, Baller JFW, Meer Hv, Lutjohann D, Kuipers F & Plosch T. (2010) The liver x-receptor gene promoter is hypermethylated in a mouse model of prenatal protein restriction. *Am J Physiol Regul Integr Comp Physiol* 298, R275–R282.
- Tang-Peronard JL, Andersen HR, Jensen TK & Heitmann BL. (2011) Endocrine-disrupting chemicals and obesity development in humans: a review. *Obes Rev* 12, 622–636.
- Tontonoz P & Spiegelman BM. (2008) Fat and beyond: the diverse biology of ppargamma. *Annu Rev Biochem* 77, 289–312.
- Valsamakis G, Anwar A, Tomlinson JW, Shackleton CH, McTernan PG, Chetty R *et al.* (2004) 11beta-hydroxysteroid dehydrogenase type 1 activity in lean and obese males with type 2 diabetes mellitus. *J Clin Endocrinol Metab* 89, 4755–4761.
- Vandenberg LN, Maffini MV, Sonnenschein C, Rubin BS & Soto AM. (2009) Bisphenol-a and the great divide: a review of controversies in the field of endocrine disruption. *Endocr Rev* 30, 75–95.
- Wade GN, Gray JM & Bartness TJ. (1985) Gonadal influences on adiposity. *Int J Obes* 9(Suppl. 1), 83–92.
- Wake DJ, Rask E, Livingstone DE, Soderberg S, Olsson T & Walker BR. (2003) Local and systemic impact of transcriptional up-regulation of 11beta-hydroxysteroid dehydrogenase type 1 in adipose tissue in human obesity. *J Clin Endocrinol Metab* 88, 3983–3988.
- Weinstock M. (2005) The potential influence of maternal stress hormones on development and mental health of the offspring. *Brain Behav Immun* 19, 296–308.
- Welberg LA, Seckl JR & Holmes MC. (2000) Inhibition of 11beta-hydroxysteroid dehydrogenase, the foeto-placental barrier to maternal glucocorticoids, permanently programs amygdala gr mrna expression and anxiety-like behaviour in the offspring. *Eur J Neurosci* 12, 1047–1054.
- Wu Q, Ohsako S, Ishimura R, Suzuki JS & Tohyama C. (2004) Exposure of mouse preimplantation embryos to 2,3,7,8-tetrachlorodibenzo-p-dioxin (tcdd) alters the methylation status of imprinted genes h19 and igf2. *Biol Reprod* 70, 1790–1797.
- Wu Q, Zhou ZJ & Ohsako S. (2006) [effect of environmental contaminants on DNA methyltransferase activity of mouse preimplantation embryos]. *Wei Sheng Yan Jiu* 35, 30–32.